Eurasian Medical Research Periodical



The Antimicrobial and Antioxidant Effects of Grapefruit (*Citrus paradise*) Peel Extract

Osama H. Abed ^{1*}		¹ Al-Rasheed University College, Dentistry Department, Baghdad,			
		Iraq			
		*Email: <u>Dr.ossama82@gmail.com</u>			
Mohammad Hussein Al-		² Iraqi Ministry of Health, Baghdad Rusafa Health Department, The			
Aubaidee ²		Specialized Dental Center in Al-maghrib Street			
Ahmed L. Azzawi ³		³ Zarcon Dental Care, Baghdad, Iraq			
Hasan N Alabbasy ⁴		⁴ Zarcon Dental Care, Baghdad, Iraq			
ABSTRACT ABSTRACT ABSTRACT ABSTRACT ABSTRACT ABSTRACT	Phytochemicals are natural compounds found in different parts of the plant (seeds, fruit, peel, rout, etc.). Phytochemicals can exhibit a therapeutic property for many health problems including cardiovascular and metabolic disorders. Grapefruit (<i>Citrus paradise</i>) is one of the citrus family that are grown in different countries around the world. The documents have shown the abundance of many phytochemicals in the grapefruit including polyphenols, flavonoids, carotenoids, and vitamins. We have aimed to prepare two extracts of the grapefruit peels, one ethanolic extract and one concentrate oil of the peels. The antimicrobial and antioxidant properties of the two extracts were experimented. The peel extract of the fruit has shown a great potential in the inhibition of bacterial strains, fungus and scavenging the free radicals. The concentrate oil extract of the grapefruit peels has shown antioxidant properties that are close to the pure ascorbic acid with maximum inhibitory effect against <i>E. coli, S. epidermidis</i> and <i>S. aureus</i> . While the antioxidant properties of the ethanolic extract of grapefruit peel were much lower than the concentrate oil or ascorbic acid, it still a good choice with a best linear association. Furthermore, ethanolic extract has shown a maximum inhibitory effect against the growth of <i>klebsiella sp.</i> and <i>C. albicans</i> which may be attributed to the characteristics of the solvent.				
Keywords:		Citrus paradise, phytochemicals, DPPH, antibacterial			

1. Introduction

The rising difficulties of antibiotic drug resistance by pathogenic organisms in recent decades has led to a continual search for novel antibiotic agents in natural plant products [1-4]. Many of these compounds are produced as secondary metabolites in plants and are frequently utilised by them to defend against microbial attack [4].

One of the most important members of the citrus genus is Citrus paradisi (Rutaceae). It's only found on the island of Barbados. Commercial grapefruit is grown in Spain, Morocco, Jordan, South Africa, Brazil, Mexico, Jamaica, and Asia [5]. Grapefruit cultivars were developed mostly in Florida and Texas, USA [6]. Citrus is a valuable fruit in the world's produce because of its nutritional worth. Citrus is a member of the Rutaceae family that originated in the subtropics and is recognized for its semisweet flavor. Numerous studies have been conducted to determine the chemical composition and antibacterial properties of essential oils extracted from the peel of various citrus species due to its nutraceutical and economic value [7].

Citrus fruits' peels are high in flavanones and polymethoxylated flavones. which are uncommon in other plants [8]. These chemicals are not only crucial for human health and the environment, but they also have a wide range of economic applications in the food as well as pharmaceutical industries. Naringin and hesperidin have a wide range of biological effects, including antioxidant, antimutagenic, analgesic, and anti-inflammatory properties. Star Ruby grapefruits (C. paradisi) and Sanguinelli oranges (C. sinensis) (Spain) were found to have antifungal properties against Penicillim digitatum [6].

Citrus essential oils have a wide range of biological actions, including antibacterial, antioxidant, anti-inflammatory, and anxiolytic [9, antimicrobial properties 10]. These capabilities have been proven to have a wide range of applications in the food business [11], for veterinary use [12, 13], human medicine [14, 15] and plants for agricultural production [16]. Our goal is to prepare two different extracts of grapefruit peels, one with ethanol as a solvent and the other without using solvent and investigate their antimicrobial and antioxidant effects.

2. Materials and Methods

2.1. Preparation of peel extracts

Iraqi grapefruit was purchased from the local market in Wasit - Iraq. The fruits were washed and cleaned perfectly, and the peel was removed and cut to small pieces. Two extracts from the peel pieces of the grapefruit were prepared. The first extract was without using any solvent. 100g of peel pieces were placed in the fruit juicer (Silvercrest, Germany) and a concentrated oil of the grape fruit peel was obtained. The other extract was prepared by adding 100g of peel pieces in 250mL 90% ethanol (Merck, Germany) and sealed the beaker with glass sealer, then the beaker was placed in the waterbath for 3h at 60 °C. The beaker was placed at the room temperature to cool dawn before opining the sealer.

2.2. Antimicrobial test

The extracts of grapefruit peels were examined against two Gram negative bacterial strains escherichia coli and klebsiella sp, and two Gram positive bacterial strains *Staphylococcus epidermidis* and *Staphylococcus aureus*, as well as one fungi (*Candida albicans*). In Petri dishes, well diffusion method was used. Two wells in the agar medium were made in a radius of 5mm, and 20μ L or 40μ L of each extract were added to the corresponded wells. The plates were incubated at 37 °C for one day, and the inhibition zones were determined in mm. *2.3. Antioxidant activity*

The activity of the grapefruit peels extracts to scavenge Diphenyl-1-picrylhydrazyl (DPPH) was determined in a spectrophotometric method [17]. A series of concentrations in methanol (Merck, Germany) of each of ascorbic acid (as a standard), concentrate grapefruit peel oil, and the ethanol grapefruit peel extract were prepared (5, 10, 20, 50, and 100 µg/mL). A weight of 0.36 g of DPPH was dissolved in 4mL methanol. 0.15mL of the DPPH solution was mixed with 3mL of each of the prepared concentrations, and with deionized water as control. The tubes were allowed to stand in dark for 30 minutes, then the absorbance of each tube was determine at 517 nm. The activity of each material was calculated from the following equation:

% Activity = (A_{DPPH} – A_{test}) / A_{DPPH}

3. Results and Discussion

3.1. Antimicrobial activity

Table 1 contain the inhibition zones that created by the grapefruit peel extracts against the growth of the strains that have been used in this study. Both extracts have shown good inhibitory effects against Gram positive and negative bacteria, as well as C. albicans. There were differences between the concentrated oil and the ethanol extract. The maximum inhibition of E. coli was obtained by the concentrated oil extract at 40µL, this was observed for S. epidermidis and S. aureus. On the other hand, ethanolic extract of grapefruit peels was shown the maximum inhibitory effect against the growth of klebsiella sp. and C. albicans. Okunowo et al. have reported that peel oil extract of grapefruit have an antimicrobial effect against the growth of several Gram positive and Gram negative bacterial strains. The authors were also reported an antifungal effects of the extract

Volume 15| December 2022

[18]. Al-Ogaili and Yasin have reported that Iraqi grapefruit peels exhibited an antimicrobial activity against the growth of *E. coli, S. epidermidis, S. aureus, klebsiella sp.,* and *C. albicans.* The authors have found that between all of the phytochemical components of the grapefruit peels, flavonoids were the most effective as antimicrobial agents [19]. Few other studies were reported an antimicrobial and antifungal activities of the grapefruits from different origins [7, 20, 21].

Type of microho	Concentrated oil extract		Ethanol extract	
Type of microbe	20µL	40µL	20µL	40µL
E. coli	10.5	14.3	5.2	7.5
<i>klebsiella</i> sp.	8.5	12.4	10.4	13.2
S. epidermidis	9.1	13.5	9.5	10
S. aureus	11.2	14.8	7.6	10.5
Candida albicans	8.6	10.1	9.2	10.8

Table 1: Inhibition zones of the grapefruit peel extracts.

3.2. Antioxidant activity

Figure 1 shows the effect of ascorbic acid to scavenge DPPH. The IC50 (the concentration of the materials that required to cause 50% inhibition of DPPH) was obtained as 30.58 μ g/mL. On the other hand, the concentrated oil extract of grapefruit peels (Figure 2) has shown to be a good antioxidant agent with an IC50 value 35.22 μ g/mL. The ethanolic extract has shown the less activity among the three tested materials, in which the IC50 was obtained as 46.70 μ g/mL.

The peels of the grapefruit has been documented to have many phytochemicals including flavonoids, carotenoids, and vitamin C and total phenolic compounds which all have an antioxidant activity [22, 23]. The grapefruit essential oil contained approximately 94% of which possess limonene an excellent antioxidant activity [24]. Several studies have reported the exhibition of good antioxidant properties from different extracts of grapefruit peels [25-28].

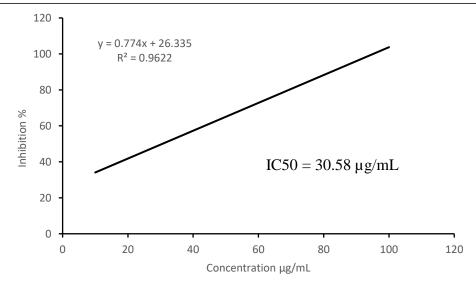


Figure 1: The antiradical activity percentage of ascorbic acid.

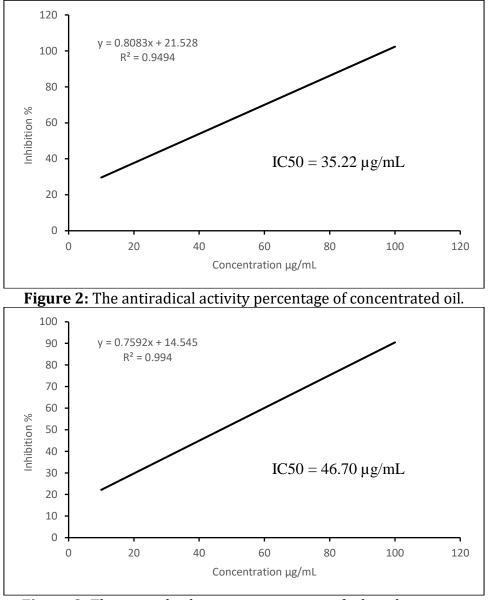


Figure 3: The antiradical activity percentage of ethanolic extract.

Conclusion

Grapefruit is full of phytochemicals with therapeutic potentials. The peel extract of the fruit has shown a great potential in the inhibition of bacterial strains, fungus and scavenging the free radicals. The concentrate oil extract of the grapefruit peels have shown antioxidant properties that are close to the pure ascorbic acid with maximum inhibitory effect against *E. coli*, *S. epidermidis* and *S. aureus*. While the antioxidant properties of the ethanolic extract of grapefruit peel were much lower than the concentrate oil or ascorbic acid, it still a good choice with a best linear association. Furthermore, ethanolic extract has shown a maximum inhibitory effects against the growth of *klebsiella sp.* and *C. albicans* which may attributed to the characteristics of the solvent.

References

- Ayoola, G., et al., Chemical analysis and antimicrobial activity of the essential oil of Syzigium aromaticum (clove). African Journal of Microbiology Research, 2008. 2(7): p. 162-166.
- 2. Belmekki, N., N. Bendimerad, and C. Bekhechi, Chemical analysis and antimicrobial activity of Teucrium polium L. essential oil from Western Algeria. Journal of Medicinal Plants Research, 2013. 7(14): p. 897-902.

- 3. Dahpour, A.A., P. Rahdari, and Z. Sobati, Chemical composition of essential oil, antibacterial activity and brine shrimp lethality of ethanol extracts from Sedum pallidum. Journal of Medicinal Plants Research, 2012. 6(16): p. 3105-3109.
- Shittu, L., et al., Antibacterial and antifungal activities of essential oils of crude extracts of Sesame radiatum against some common pathogenic microorganisms. Iranian Journal of pharmacology and Therapeutics, 2007. 6(2): p. 165-0.
- Salman, A. T., Ismail, A. H., Rheima, A. M., Abd, A. N., Habubi, N. F., & Abbas, Z. S. (2021, March). Nano-Synthesis, characterization and spectroscopic Studies of chromium (III) complex derived from new quinoline-2-one for solar cell fabrication. In *Journal of Physics: Conference Series* (Vol. 1853, No. 1, p. 012021). IOP Publishing
- 6. Ortuño, A., et al., Citrus paradisi and Citrus sinensis flavonoids: Their influence in the defence mechanism against Penicillium digitatum. Food chemistry, 2006. 98(2): p. 351-358.
- 7. Vasek, O., et al., Antibacterial activity of Citrus paradisi essential oil. Journal of Natural Products, 2015. 8: p. 16-26.
- Ahmed, H., Hussein, S. N., Ali, R. A., Almashhadani, H. A., & Ayvaz, A. (2022). Environmental effects on intestinal parasitic disease transmission in Mosul governorate. Journal of Pharmaceutical Negative Results¦ Volume, 13(3), 269.
- 9. Zia-ur-Rehman, Citrus peel extract-A natural source of antioxidant. Food Chemistry, 2006. 99(3): p. 450-454.
- 10. Chutia, M., et al., Antifungal activity and chemical composition of Citrus reticulata Blanco essential oil against phytopathogens from North East India. LWT-Food Science and Technology, 2009. 42(3): p. 777-780.
- 11. Hameed, R. Y., Nathir, I., Abdulsahib, W. K., & Almashhadani, H. A. (2022). Study the effect of biosynthesized gold nanoparticles on the enzymatic activity of alpha-Amylase. Research Journal of

Pharmacy and Technology, 15(8), 3459-3465.

- 12. Imran, K., et al., Extraction and applications of Grapefruit (Citrus paradise) peel oil against E. coli. Pakistan Journal of Nutrition, 2013. 12(6): p. 534.
- 13. Rheima, A. M., Anber, A. A., Abdullah, H. I., & Ismail, A. H. (2021). Synthesis of Alpha-Gamma Aluminum Oxide Nanocomposite via Electrochemical Method for Antibacterial Activity. *Nano Biomed. Eng*, *13*(1), 1-5.
- 14. Kadhim, M. (2022). Total Oxidants, Lipid Peroxidation and Antioxidant Capacity in the Serum of Rheumatoid Arthritis Patients. Journal of Pharmaceutical Negative Results¦ Volume, 13(3).
- 15. Roussenova, N., Antibacterial activity of essential oils against the etiological agent of American foulbrood disease (Paenibacillus larvae). Bulgarian Journal of Veterinary Medicine, 2011. 14(1): p. 17-24.
- 16. Ahmed, G. S., Shari, F. H., Alwan, H. A., Obaid, R. F., Almashhadani, H. A., & Kadhim, M. M. (2022). The Level of Nitric Oxide Synthase and Nitric Oxide in Hypertensive Women. Journal of Pharmaceutical Negative Results¦ Volume, 13(3), 237.
- 17. Yin, X., C.L. Gyles, and J. Gong, Grapefruit juice and its constituents augment the effect of low pH on inhibition of survival and adherence to intestinal epithelial cells of Salmonella enterica serovar Typhimurium PT193. International Journal of Food Microbiology, 2012. 158(3): p. 232-238.
- Oliveira, S.A.C., et al., The antimicrobial effects of Citrus limonum and Citrus aurantium essential oils on multi-species biofilms. Brazilian Oral Research, 2013. 28: p. 22-27.
- 19. Mahdi, A., Abbas, Z. S., Hassanain, K., & d Ha, I. (2020). Synthesis, characterization, spectroscopic, and biological activity studies of Nano scale Zn (II), Mn (II) and Fe (II) theophylline complexes.
- 20. Mahdi, M., et al. Green synthesis of gold NPs by using dragon fruit: Toxicity and

wound healing. in Journal of Physics: Conference Series. 2021. IOP Publishing.

- 21. Okunowo, W.O., et al., Essential oil of grape fruit (Citrus paradisi) peels and its antimicrobial activities. 2013.
- 22. Al-Ogaili, N.A. and Z. Yasin, Qualitative and Quantitative Investigation of Iraqi Grapefruit (Citrus padisi) Flavonoids From Peel and Seeds and Comparing Their Aqueous Extracts for Antimicrobial Activity. Iraqi Journal of Science, 2016: p. 2627-2633.
- 23. Gupta, V., et al., Pharmacological potentials of Citrus paradisi-an overview. Int J Phytother Res, 2011. 1(1): p. 8-17.
- 24. Kirbaşlar, F.G., et al., Antimicrobial activity of Turkish citrus peel oils. Pak. J. Bot, 2009. 41(6): p. 3207-3212.
- 25. Aziz, S. N., Al Marjani, M. F., Rheima, A. M., & Al Kadmy, I. M. (2022). Antibacterial, antibiofilm, and antipersister cells formation of green synthesis silver nanoparticles and graphene nanosheets against Klebsiella pneumoniae. *Reviews* in Medical Microbiology, 33(1), 56-63.
- 26. Uckoo, R.M., et al., Grapefruit (Citrus paradisi Macfad) phytochemicals composition is modulated by household processing techniques. Journal of food science, 2012. 77(9): p. C921-C926.
- 27. Misharina, T.A. and A.L. Samusenko, Antioxidant properties of essential oils from lemon, grapefruit, coriander, clove, and their mixtures. Applied Biochemistry and Microbiology, 2008. 44(4): p. 438-442.
- 28. Castro-Vazquez, L., et al., Bioactive flavonoids, antioxidant behaviour, and cytoprotective effects of dried grapefruit peels (Citrus paradisi Macf.). Oxidative Medicine and Cellular Longevity, 2016. 2016.
- 29. Al Marjani, M., Aziz, S. N., Rheima, A. M., & Abbas, Z. S. (2021). Impact of Chromium Ooxide Nnanoparticles on gGrowth and bBiofilm fFormation of pPersistence Klebsiella pneumoniae iIsolates. *Nano Biomed. Eng*, *13*(3), 321-327.

- Mohammed, A. K., Al-Shaheeb, S., Fawzi, O. F., Almashhadani, H. A., & Kadhim, M. M. (2022). Evaluation of Interlukein-6 and Vitamin D in Patients with COVID-19. Research Journal of Biotechnology Vol, 17(10).
- 31. Gorinstein, S., et al., Changes in plasma lipid and antioxidant activity in rats as a result of naringin and red grapefruit supplementation. Journal of agricultural and food chemistry, 2005. 53(8): p. 3223-3228.
- 32. Oboh, G. and A. Ademosun, Characterization of the antioxidant properties of phenolic extracts from some citrus peels. Journal of food science and technology, 2012. 49(6): p. 729-736.