

Pseudomonas aeruginosa and the multifactorial antibiotic resistance

Sarah Ahmed Hasan Microbiologist, University of Kirkuk, Kirkuk, Iraq. Author email: sarahahmed100@uokirkuk.edu.iq Pseudomonas aeruginosa is a gram negative bacterium, causes many infections specially in immunocompromised individuals and causes critical infections in cystic fibrosis patients. It has remarkable ability to resist various kinds of antibiotics by intrinsic, acquired and adaptive resistance mechanisms, because of the diversity in resistance mechanisms this bacterium contributes in developing multidrug-resistant strains. P.aeruginosa infections' treatment is considered as a significant challenge which need to develop new therapies to treat these infections **Keywords:** Pseudomonas multifactorial aeruginosa, antibiotic resistance,

Introduction

Pseudomonas aeruginosa is an opportunistic bacterium ,infects the immunocompromised individuals and causes critical infections in cystic fibrosis patients, this bacterium has a great ablility to survive in a broad range of environments[1].

P.aeruginosa infections' treatment is considered as significant challenge cause of the bacterium ability to resist various kinds of antibiotics[2-7]. Carbapenem-resistant *P. aeruginosa* is recently listed by the World Health Organization as a critical problem which need to develop new therapies to treat these infections[8]. Over use and/or misuse of antibiotics lead to develop multidrug-resistant strains of *P. aeruginosa*[2-7].

This study focused on the recent multifactorial antibiotic resistance mechanisms (intrinsic , acquired and adaptive) in *P. aeruginosa* .

1- Intrinsic antibiotic resistance

Define as the bacterial innate ability to minimize the antibiotic efficacy by inherent functional or structural characteristics[9].

Pseudomonas aeruginosa has a great scale of this kind of antibiotic resistance by various mechanisms[10] as mentioned in (Fig. 1)[11].



Figure 1- Antibiotic resistant mechanisms

1.1. Outer membrane permeability

P. aeruginosa has selective barrier outer membrane against penetration of antibiotics, contains porins, generally, the porins' family consist of four classes as mentioned in Fig.2. In *P. aeruginosa*, there are important and specific porins act as this kind of antibiotic resistance as mentioned in Fig.3[12].

The lower permeability of *P. aeruginosa* outer membrane compared with other bacteria is due

to the closed OprF channels. While, the absence of OprF in this bacterium caused increasing in biofilm formation[13].

The absence of OprD in this bacterium increases the resistance against carbapenem antibiotics and the overexpression of the smallest porin (OprH) because of Mg2+ starvation is associated with the increasing of resistance against gentamicin and polymyxin B[14].



Figure 2- The four classes of porins family



Figure 3- The porins of Pseudomonas aeruginosa

1.2. Efflux systems

Expelling toxic compounds out of the bacterial cells, consist of five families [15] as mentioned in Fig.4. In *P. aeruginosa* ,the proteins of RND

efflux pumps family play a significant role in their antimicrobial resistance[16].

This bacterium has twelve RND family efflux pumps, four of them contribute in antimicrobial resistance[17] as mentioned in Fig.5.





Figure 5- Four RND family efflux pumps in Pseudomonas aeruginosa

1-3 Antibiotic-inactivating enzymes

These enzymes can modify or break down antibiotics. Esters and amides chemical bonds in many antibiotics are broken down by these enzymes such as aminoglycoside-modifying enzymes and β -lactamases which commonly produced by *P.aeruginosa*[18,19].

This bacterium has an inducible ampC gene, like other Gram-negative bacteria, this gene encoding for production of β -lactamase which can hydrolysis the amide bond of β -lactam ring [18]. There are four classes of β - lactamases: A, B, C and D, classified by depending on the sequences of amino acids[20].

Some stains *P. aeruginosa* can produce extended-spectrum- β -lactamases (ESBLs) which give a high level of resistance against various classes of β -lactam antimicrobials. Clavulanate, tazobactam and sulbactam considered as β -lactamase inhibitors used to overcome β -lactamase enzymes and useful in combination therapies[21].

2. Acquired antibiotic resistance

This kind of resistance consists of two ways, detailed below.

2.1. Resistance by mutations

Mutations cause decreasing of antibiotic uptaking, overexpression of efflux systems, inactivating enzymes and modifications of antibiotic targets, thus the bacteria can survive against antibiotics attack[22].

Such as, a deficiency in OprD cause a high degree of resistance in *P. aeruginosa* against

carbapenem antibiotics, specifically imipenem[23].

2.2. Acquisition of resistance genes

Acquisition of plasmids, prophages, integrons and transposons via horizontal gene transfer causes this kind of resistance[10] as mentioned in Fig.6&7. In *P. aeruginosa* acquisition of resistance genes against aminoglycosides and β lactams has been conducted[24].



Figure 6- Acquisition of resistance genes



Figure 7- Horizontal gene transfer mechanisms

3- Adaptive antibiotic resistance

Another mechanism to increase the bacterial ability to survive from antibiotic concentrations[25,26].

3-1 Biofilm-mediated resistance

This kind of mechanism makes the bacterial cells less sensitive to antibiotics and immune

response of the host[27], mentioned in Fig 8[28].

Two-component regulatory systems regulate the formation of biofilms; called by GacS/GacA & RetS/LadS, exopolysaccharides & cdi- GMP in *P. aeruginosa* [29]. This bacterium has three main quorum sensing as mentioned in Fig 9 contribute in the formation of biofilms[11,30].





Figure 9- Quorum sensing systems in Pseudomonas aeruginosa

3.2.Bacterial persister cells

Another resistant mechanism against antibiotics with many significant properties as mentioned in Fig.10 [11].



potential to become multidrug-tolerant.

Figure 10- Properties of persister cells in Pseudomonas aeruginosa

Conclusion

The treatment of infections that caused by *P. aeruginosa* considered as a serious challenge. *P. aeruginosa* has multifactorial antibiotic resistance because of the diversity in resistance mechanisms (intrinsic, acquired and adaptive) this bacterium contributes in developing multidrug-resistant strains.

In addition to that *P. aeruginosa* able to form biofilms and persister cells which are the causative of continuous and difficult infections in Cystic fibrosis patients.

Reference

 Silby, M.W., Winstanley, C., Godfrey, S.A., Levy, S.B., Jackson, R.W., 2011. Pseudomonas genomes: diverse and adaptable. FEMS Microbiol Rev 35, 652– 680.

- 2- Hasan SA, Abass KS. Prevalence of Gram Negative Bacteria Isolated from Patients with Burn Infection and their Antimicrobial Susceptibility Patterns in Kirkuk City, Iraq. Indian J Public Health Res Dev 2019;10(8).
- 3- Hasan SA, Najati AM, Abass KS. Prevalence and antibiotic resistance of "pseudomonas aeruginosa" isolated from clinical samples in Kirkuk City, Iraq. Eurasia J Biosci. 2020;14(1):1821-5.
- 4- Hasan SA, Najati AM, Abass KS. Isolation and identification of multidrug resistant "Pseudomonas aeruginosa" from burn wound infection in Kirkuk City. Eurasia J Biosci 2019;13:1045-50.
- 5- Ahmed Hasan S, Fakhraddin Raheem T, Mohammed Abdulla H. Phenotypic,

Antibiotyping, and Molecular Detection of Klebsiella Pneumoniae Isolates from Clinical Specimens in Kirkuk, Iraq. Arch Razi Inst. 2021 Oct 31;76(4):1061-1067. doi: 10.22092/ari.2021.355770.1721. PMID: 35096342; PMCID: PMC8790972.

- 6- Ahmed Hasan, S., Mohammed Bakr, M. (2022). Bacteriological and Molecular Detection of Klebsiella oxytoca and its Resistance to Antibiotics among Clinical Specimens from Kirkuk, Iraq. Archives of Razi Institute, 77(5), 1521-1525. doi: 10.22092/ari.2022.357753.209
- 7- Hasan SA, Saleh I, Ali H. Bacteriological and Molecular Detection of Staphylococcus Aureus and its Resistance to Methicillin among Specimens from Kirkuk Community. Ann Rom Soc Cell Biol. 2021;25(7):461-73
- 8- Tacconelli, E., Magrini, N., Carmeli, Y., Harbarth, S., Kahlmeter, G., Kluytmans, J., Mendelson, M., Pulcini, C., Singh, N., Theuretzbacher, U., 2017. Global priority list of antibiotic-resistant bacteria to guide research, discovery, and development of new antibiotics. World Health Organization 1-7(http://www.who.int/medicines/ publications/WHO-PPL-Short_Summary_25Feb-ET_NM_WHO.pdf. Last date of access: Nov.18.2018).
- 9- Blair, J.M., Webber, M.A., Baylay, A.J., Ogbolu, D.O., Piddock, L.J., 2015. Molecular mechanisms of antibiotic resistance. Nat Rev Microbiol 13, 42–51.
- 10-Breidenstein, E.B., de la Fuente-Nunez, C., Hancock, R.E., 2011. Pseudomonas aeruginosa: all roads lead to resistance. Trends Microbiol 19, 419–426.
- 11-Pang Z, Raudonis R, Glick BR, Lin TJ, Cheng Z. Antibiotic resistance in Pseudomonas aeruginosa: mechanisms and alternative therapeutic strategies. Biotechnol Adv. 2019 Jan-Feb;37(1):177-192. doi: 10.1016/j.biotechadv.2018.11.013. Epub 2018 Nov 27. PMID: 30500353.

- 12-Hancock, R.E., Brinkman, F.S., 2002. Function of pseudomonas porins in uptake and efflux. Annu Rev Microbiol 56, 17–38.
- 13-Bouffartigues, E., Moscoso, J.A., Duchesne, R., Rosay, T., Fito-Boncompte, L., et al., 2015. The absence of the Pseudomonas aeruginosa OprF protein leads to increased biofilm formation through variation in c-di-GMP level. Front Microbiol 6, 630.
- 14-Macfarlane, E.L., Kwasnicka, A., Ochs, M.M., Hancock, R.E., 1999. PhoP-PhoQ homologues in Pseudomonas aeruginosa regulate expression of the outermembrane protein OprH and polymyxin B resistance. Mol Microbiol 34, 305–316.
- 15-Sun, J., Deng, Z., Yan, A., 2014. Bacterial multidrug efflux pumps: mechanisms, physiology and pharmacological exploitations. Biochem Biophys Res Commun 453, 254–267.
- 16-Li, X.Z., Nikaido, H., 2009. Effluxmediated drug resistance in bacteria: an update. Drugs 69, 1555–1623.
- 17-Dreier, J., Ruggerone, P., 2015. Interaction of antibacterial compounds with RND e ffl ux pumps in Pseudomonas aeruginosa. Front Microbiol 6, 660.
- 18-Wright, G.D., 2005. Bacterial resistance to antibiotics: enzymatic degradation and modification. Adv Drug Deliv Rev 57, 1451–1470.
- 19-Wolter, D.J., Lister, P.D., 2013. Mechanisms of beta-lactam resistance among Pseudomonas aeruginosa. Curr Pharm Des 19, 209–222.
- 20-Bush, K., Jacoby, G.A., 2010. Updated functional classification of betalactamases. Antimicrob Agents Chemother 54, 969–976.
- 21-Rawat, D., Nair, D., 2010. Extendedspectrum beta-lactamases in Gram negative bacteria. J Glob Infect Dis 2, 263–274.
- 22-Munita, J.M., Arias, C.A., 2016. Mechanisms of antibiotic resistance. Microbiol Spectr 4. Murphy, T.F., 2009. Pseudomonas aeruginosa in adults with

chronic obstructive pulmonary disease. Curr Opin Pulm Med 15, 138–142.

- 23-Fang, Z.L., Zhang, L.Y., Huang, Y.M., Qing, Y., Cao, K.Y., et al., 2014. OprD mutations and inactivation in imipenem-resistant Pseudomonas aeruginosa isolates from China. Infect Genet Evol 21, 124–128.
- 24-Hong, D.J., Bae, I.K., Jang, I.H., Jeong, S.H., Kang, H.K., Lee, K., 2015. Epidemiology and characteristics of Metallo-beta-Lactamase-producing Pseudomonas aeruginosa. Infect Chemother 47, 81–97.
- 25-Sandoval-Motta, S., Aldana, M., 2016. Adaptive resistance to antibiotics in bacteria: a systems biology perspective. Wiley Interdiscip Rev Syst Biol Med 8, 253–267.
- 26-Taylor, P.K., Yeung, A.T., Hancock, R.E., 2014. Antibiotic resistance in Pseudomonas aeruginosa biofilms: towards the development of novel antibiofilm therapies. J Biotechnol 191, 121– 130.

Rasamiravaka, T., Labtani, Q., Duez, P., El Jaziri, M., 2015. The formation of biofilms by

- 27-Muhammad MH, Idris AL, Fan X, Guo Y, Yu Y, Jin X, Qiu J, Guan X, Huang T. Beyond Risk: Bacterial Biofilms and Their Regulating Approaches. Front Microbiol.
 2020 May 21;11:928. doi: 10.3389/fmicb.2020.00928. PMID: 32508772; PMCID: PMC7253578.
- 28-Pseudomonas aeruginosa: a review of the natural and synthetic compounds interfering with control mechanisms. Biomed Res Int 2015, 759348.
- 29-Kang, D., Turner, K.E., Kirienko, N.V., 2017. PqsA promotes Pyoverdine production via biofilm formation. Pathogens 7. Karaiskos, I., Souli, M., Giamarellou, H.